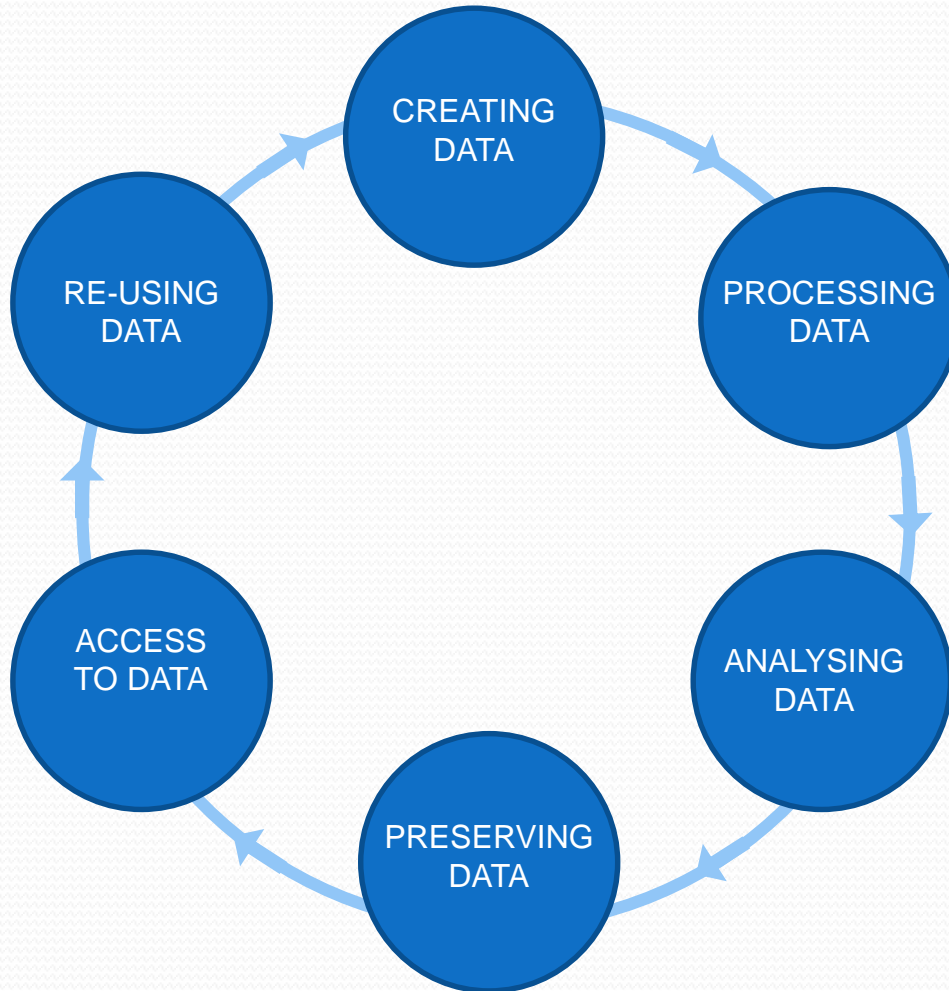


Tipping the balance

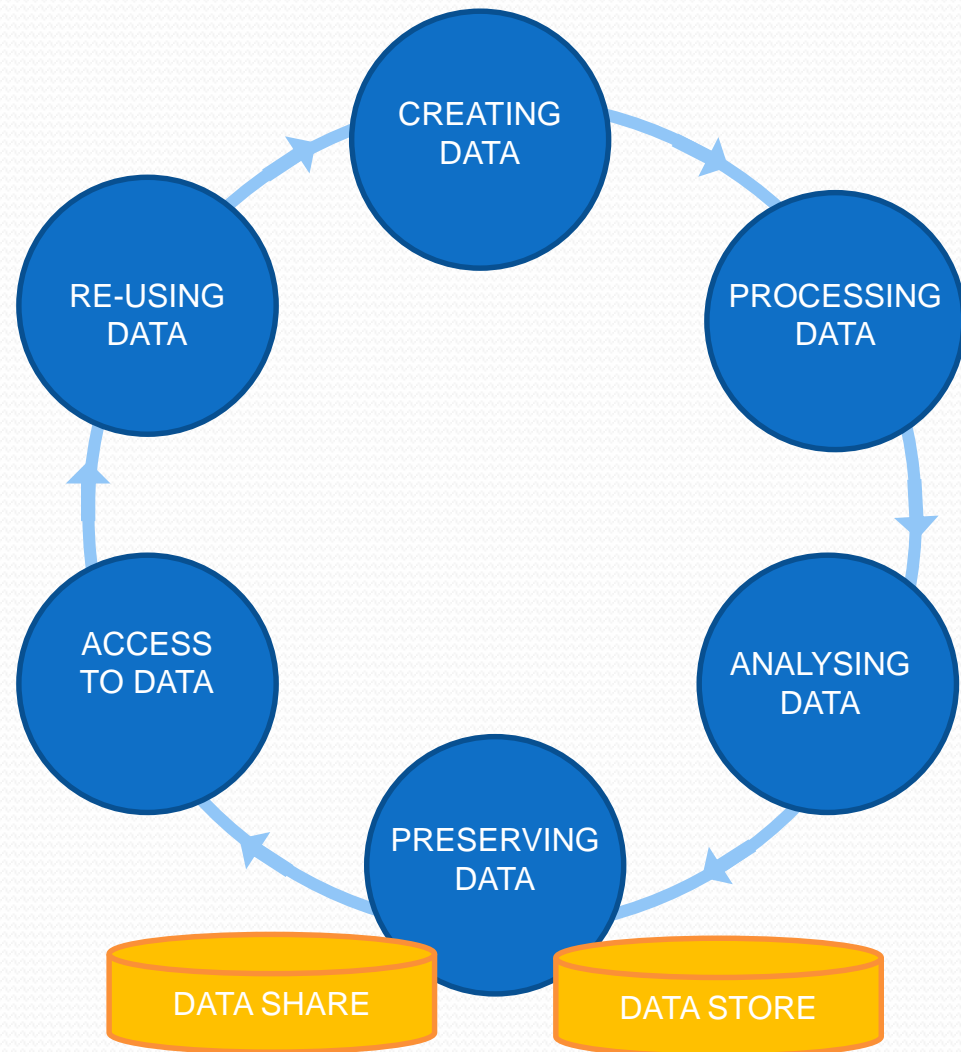
**introducing data management
on a centre-wide level**

Tomasz Zieliński, Eilidh Troup, Andrew Millar

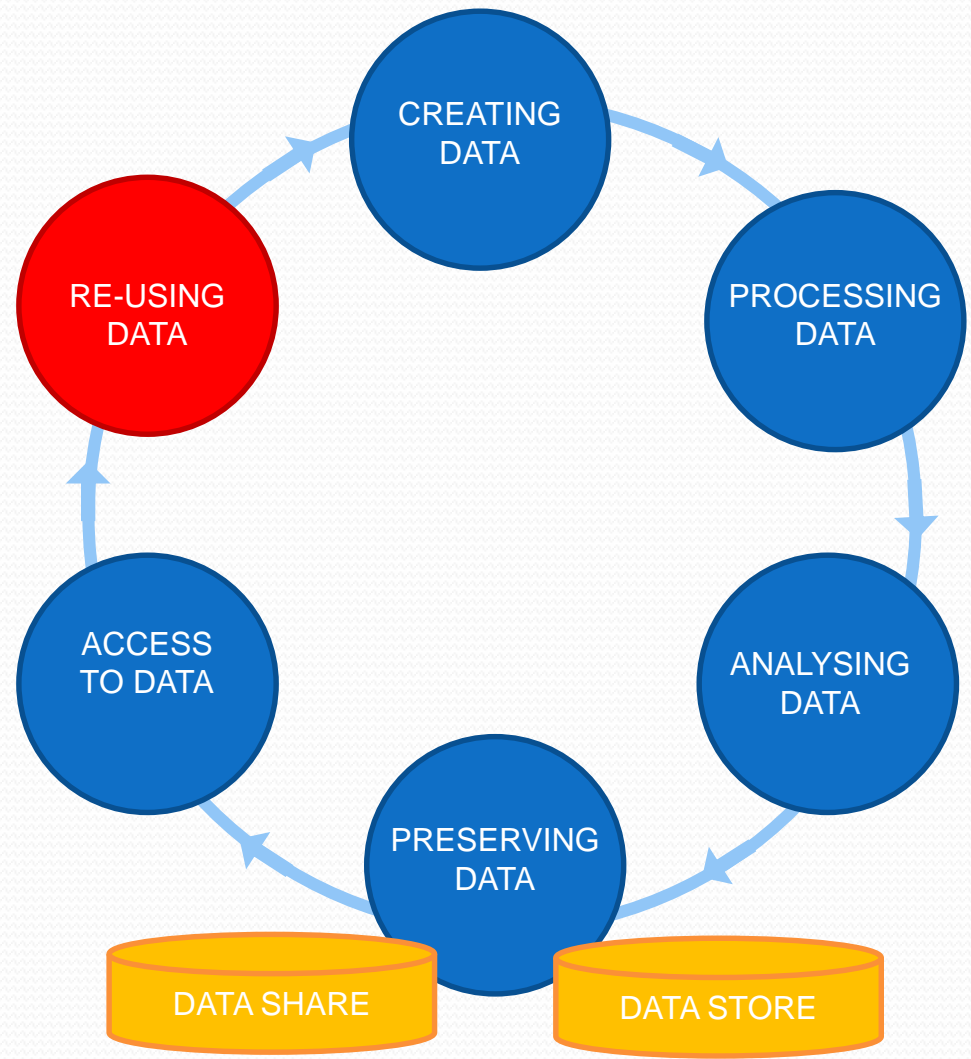
Data life-cycle



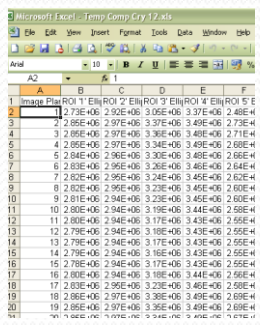
Data life-cycle



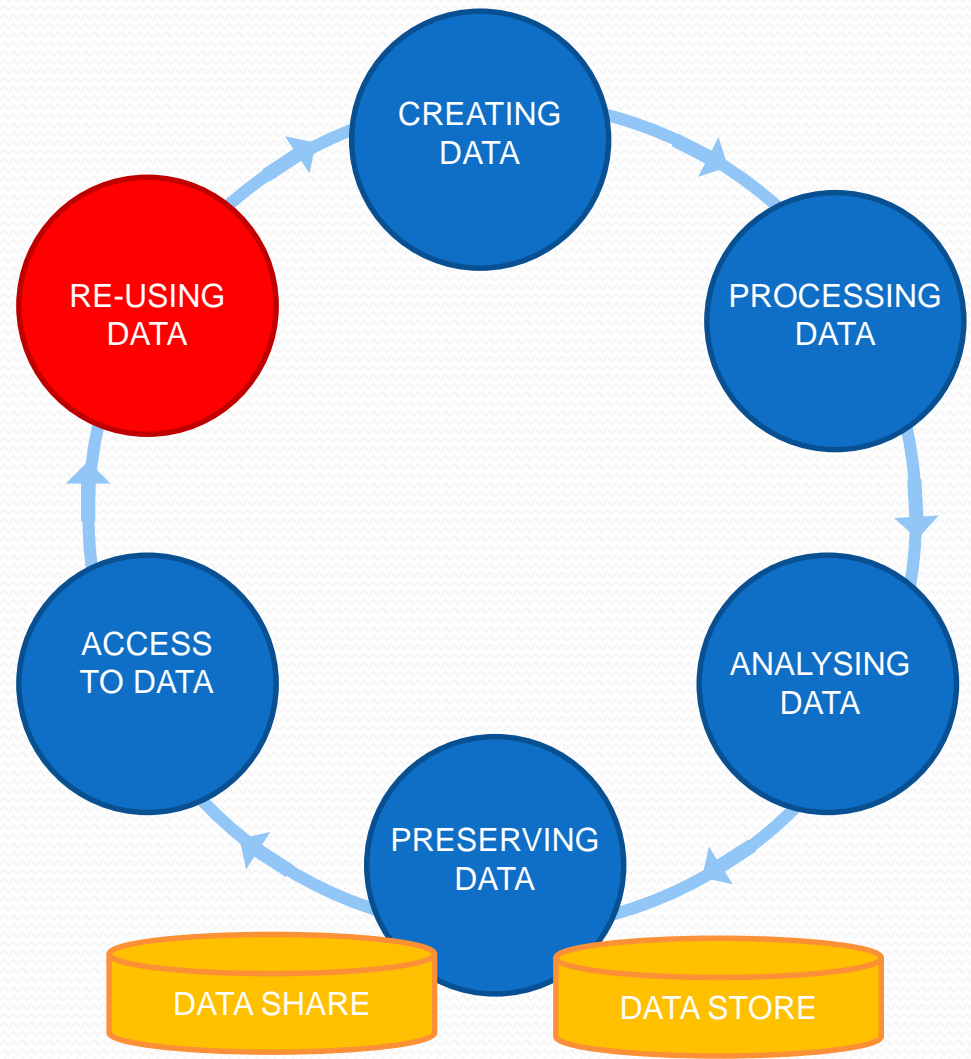
Data life-cycle



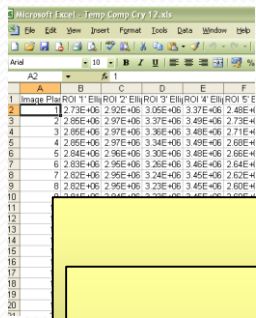
Data life-cycle



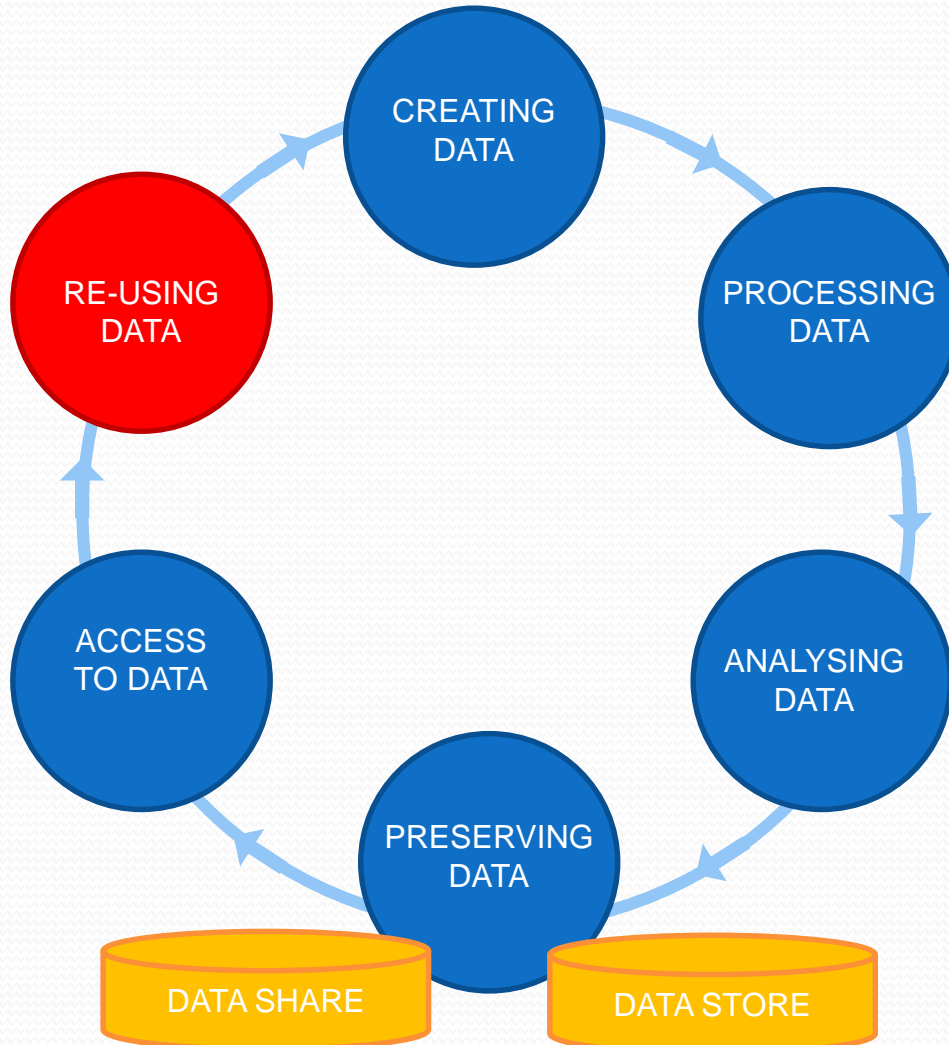
	A	B	C	D	E	F
1	Image	ROI 1	ROI 2	ROI 3	ROI 4	ROI 5
2		2.73E+06	2.92E+06	3.06E+06	3.37E+06	2.48E+06
3		2.89E+06	2.97E+06	3.37E+06	3.49E+06	2.73E+06
4		3.28E+06	2.97E+06	3.36E+06	3.48E+06	2.71E+06
5		4.28E+06	2.97E+06	3.34E+06	3.49E+06	2.68E+06
6		5.04E+06	2.96E+06	3.36E+06	3.49E+06	2.66E+06
7		6.28E+06	2.95E+06	3.36E+06	3.46E+06	2.64E+06
8		7.28E+06	2.95E+06	3.24E+06	3.45E+06	2.62E+06
9		8.03E+06	2.95E+06	3.22E+06	3.45E+06	2.60E+06
10		9.281E+06	2.94E+06	3.23E+06	3.45E+06	2.60E+06
11		10.280E+06	2.94E+06	3.19E+06	3.44E+06	2.58E+06
12		11.280E+06	2.94E+06	3.17E+06	3.43E+06	2.56E+06
13		12.279E+06	2.94E+06	3.18E+06	3.43E+06	2.55E+06
14		13.279E+06	2.94E+06	3.17E+06	3.43E+06	2.55E+06
15		14.279E+06	2.94E+06	3.18E+06	3.43E+06	2.55E+06
16		15.278E+06	2.94E+06	3.17E+06	3.43E+06	2.55E+06
17		16.280E+06	2.94E+06	3.18E+06	3.44E+06	2.56E+06
18		17.280E+06	2.95E+06	3.22E+06	3.46E+06	2.59E+06
19		18.289E+06	2.97E+06	3.30E+06	3.49E+06	2.69E+06
20		19.289E+06	2.97E+06	3.35E+06	3.49E+06	2.69E+06



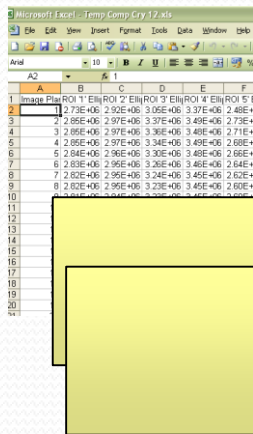
Data life-cycle



	A	B	C	D	E	F
1	Image	ROI 1	ROI 2	ROI 3	ROI 4	ROI 5
2		2.73E+06	2.92E+06	3.06E+06	3.37E+06	2.48E+H
3		2.89E+06	2.97E+06	3.37E+06	3.49E+06	2.75E+H
4		2.89E+06	2.97E+06	3.36E+06	3.48E+06	2.71E+H
5		2.89E+06	2.97E+06	3.34E+06	3.49E+06	2.68E+H
6		2.94E+06	2.96E+06	3.36E+06	3.49E+06	2.66E+H
7		2.83E+06	2.95E+06	3.36E+06	3.46E+06	2.64E+H
8		2.82E+06	2.95E+06	3.24E+06	3.45E+06	2.62E+H
9		2.82E+06	2.95E+06	3.23E+06	3.45E+06	2.60E+H
10						
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17						
18						
19						
20						
21						



Data life-cycle



	A	B	C	D	E	F
1	Image Plot	EukROI 1	EukROI 2	EukROI 3	EukROI 4	EukROI 5
2		2.73E+06	2.92E+06	3.05E+06	3.37E+06	2.48E+06
3		2.88E+06	2.97E+06	3.37E+06	3.48E+06	2.75E+06
4		2.89E+06	2.97E+06	3.36E+06	3.48E+06	2.71E+06
5		4.28E+06	2.97E+06	3.34E+06	3.49E+06	2.68E+06
6		5.04E+06	2.96E+06	3.38E+06	3.48E+06	2.66E+06
7		6.283E+06	2.95E+06	3.36E+06	3.46E+06	2.64E+06
8		7.282E+06	2.95E+06	3.24E+06	3.45E+06	2.62E+06
9		8.282E+06	2.95E+06	3.23E+06	3.45E+06	2.60E+06
10						
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16						
17						
18						
19						
20						

Data Creator: Martin, Sarah F.; Noordally, Zeenat B.; van Ooijen, Gerben; Barrios-Llerena, Martin E.; Simpson, T. Ian; Millar, Andrew J.; Hindle, Matthew M.; Thierry Le Bihan

Date Available: 2014-06-24

Citation: Martin, Sarah F.; Noordally, Zeenat B.; van Ooijen, Gerben; Barrios-Llerena, Martin E.; Simpson, T. Ian; Millar, Andrew J.; Hindle, Matthew M.; Thierry Le Bihan. (2014). The reduced kinome of *Ostreococcus tauri*: core eukaryotic signalling components in a tractable model species, [Dataset]. University of Edinburgh. SynthSys and School of Biological Sciences. <http://dx.doi.org/10.7488/ids/72>.

Dataset Description (abstract):

Background The current knowledge of eukaryote signalling originates from phenotypically diverse organisms. There is a pressing need to identify conserved signalling components among eukaryotes, which will lead to the transfer of knowledge across kingdoms. Two useful properties of a eukaryote model for signalling are (1) reduced signalling complexity, (2) conservation of signalling components. The alga *Ostreococcus tauri* is described as the smallest free-living eukaryote. With less than 8,000 genes, it represents a highly constrained genomic palette. **Results** Our survey revealed 133 protein kinases and 34 protein phosphatases (1.7% and 0.4% of the proteome). We conducted phospho-proteomic experiments and constructed domain structures and phylogenies for the catalytic protein-kinases. For each of the major kinases families we review the completeness and divergence of *O. tauri* representatives in comparison to the well-studied kinomes of the laboratory models *Arabidopsis thaliana* and *Saccharomyces cerevisiae*, and of *Homo sapiens*. Many kinase clades in *O. tauri* were reduced to a single member, in preference to the loss of family diversity, whereas TKL and ABC1 clades were expanded. We also identified kinases that have been lost in *A. thaliana* but retained in *O. tauri*. For three, contrasting eukaryotic pathways – TOR, MAPK, and the circadian clock – we established the subset of conserved components and demonstrate conserved sites of substrate phosphorylation and kinase motifs. **Conclusions** We conclude that *O. tauri* satisfies our two central requirements. Several of its kinases are more closely related to *H. sapiens* orthologs than *S. cerevisiae* is to *H. sapiens*. The greatly reduced kinome of *O. tauri* is therefore a suitable model for signalling in free-living eukaryotes.

DATA SHARE

DATA STORE

RESEARCH ARTICLE

Open Access

The reduced kinome of *Ostreococcus tauri*: core eukaryotic signalling components in a tractable model species

Matthew M Hindle^{1,4}, Sarah F Martin^{1,2}, Zeenat B Noordally^{1,2}, Gerben van Ooijen^{1,2}, Martin E Barrios-Llerena^{1,2}, T Ian Simpson^{3,4}, Thierry Le Bihan^{1,2} and Andrew J Millar^{1,2*}

Abstract

Background: The current knowledge of eukaryote signalling originates from phenotypically diverse organisms. There is a pressing need to identify conserved signalling components among eukaryotes, which will lead to the transfer of knowledge across kingdoms. Two useful properties of a eukaryote model for signalling are (1) reduced signalling complexity, and (2) conservation of signalling components. The alga *Ostreococcus tauri* is described as the smallest free-living eukaryote. With less than 8,000 genes, it represents a highly constrained genomic palette.

Results: Our survey revealed 133 protein kinases and 34 protein phosphatases (1.7% and 0.4% of the proteome). We conducted phosphoproteomic experiments and constructed domain structures and phylogenies for the catalytic protein-kinases. For each of the major kinase families we review the completeness and divergence of *O. tauri* representatives in comparison to the well-studied kinomes of the laboratory models *Arabidopsis thaliana* and *Saccharomyces cerevisiae*, and of *Homo sapiens*. Many kinase clades in *O. tauri* were reduced to a single member, in preference to the loss of family diversity, whereas TKL and ABC1 clades were expanded. We also identified kinases that have been lost in *A. thaliana* but retained in *O. tauri*. For three, contrasting eukaryotic pathways – TOR, MAPK, and the circadian clock – we established the subset of conserved components and demonstrate conserved sites of substrate phosphorylation and kinase motifs.

Conclusions: We conclude that *O. tauri* satisfies our two central requirements. Several of its kinases are more closely related to *H. sapiens* orthologs than *S. cerevisiae* is to *H. sapiens*. The greatly reduced kinome of *O. tauri* is therefore a suitable model for signalling in free-living eukaryotes.

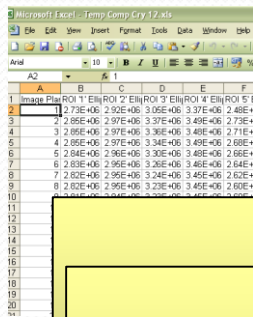
Keywords: Conserved eukaryote signalling, Protein kinase phylogeny, *Ostreococcus tauri*, Model kinome, Phosphorylation, TOR signalling, MAPK cascade, Circadian clock

Background

Protein kinases are a major component of the complex signalling networks that coordinate all fundamental cellular processes, including transcription, cell cycle and metabolism. Protein kinases and phosphatases elicit reversible phosphorylation, which enable the rapid cellular responses that are crucial for survival in a continually changing environment. Protein kinases

activate and deactivate proteins by addition of the gamma-phosphate from ATP to serine (S), threonine (T), tyrosine (Y), aspartate (D) or histidine (H) amino acid residues [1]. Cascades of consecutive kinase-mediated phosphorylation events constitute the backbone of signalling pathways [2]. The complexity of the signalling networks scales with size. Part of this complexity is constrained by the number of genes encoding protein kinases, also known as the kinome. The number of encoded protein kinases in free-living eukaryotes ranges from as little as 126 kinases in *Saccharomyces cerevisiae* [3] to ~1000

* Correspondence: andrew.millar@ed.ac.uk
¹SynthSys and School of Biological Sciences, University of Edinburgh, Edinburgh EH9 3FD UK



Data Creator: Martin, Sarah F.; Noordally, Zeenat B.; Millar, Andrew J.; Hindle, Matthew M.

Date Available: 2014-06-24

Citation: Martin, Sarah F.; Noordally, Zeenat B.; Millar, Andrew J.; Hindle, Matthew M.; Simpson, T Ian; Barrios-Llerena, Martin E.; Le Bihan, Thierry; Ooijen, Gerben van; et al. *Ostreococcus tauri*: core eukaryotic signalling components in a tractable model species. *BMC Genomics* 2014, **15**:640. doi:10.1186/s12864-014-0640-7

Dataset Description (abstract):

Background The current knowledge of eukaryote signalling originates from phenotypically diverse organisms. There is a pressing need to identify conserved signalling components among eukaryotes, which will lead to the transfer of knowledge across kingdoms. Two useful properties of a eukaryote model for signalling are (1) reduced signalling complexity, and (2) conservation of signalling components. The alga *Ostreococcus tauri* is described as the smallest free-living eukaryote. With less than 8,000 genes, it represents a highly constrained genomic palette. We conducted phosphoproteomic experiments and constructed domain structures and phylogenies for the catalytic protein-kinases. For each of the major kinase families we review the completeness and divergence of *O. tauri* representatives in comparison to the well-studied kinomes of the laboratory models *Arabidopsis thaliana* and *Saccharomyces cerevisiae*, and of *Homo sapiens*. Many kinase clades in *O. tauri* were reduced to a single member, in preference to the loss of family diversity, whereas TKL and ABC1 clades were expanded. We also identified kinases that have been lost in *A. thaliana* but retained in *O. tauri*. For three, contrasting eukaryotic pathways – TOR, MAPK, and the circadian clock – we established the subset of conserved components and demonstrate conserved sites of substrate phosphorylation and kinase motifs. We conclude that *O. tauri* satisfies our two central requirements. Several of its kinases are more closely related to *H. sapiens* orthologs than *S. cerevisiae* is to *H. sapiens*. The greatly reduced kinome of *O. tauri* is therefore a suitable model for signalling in free-living eukaryotes.

DATA SHARE

Getting metadata

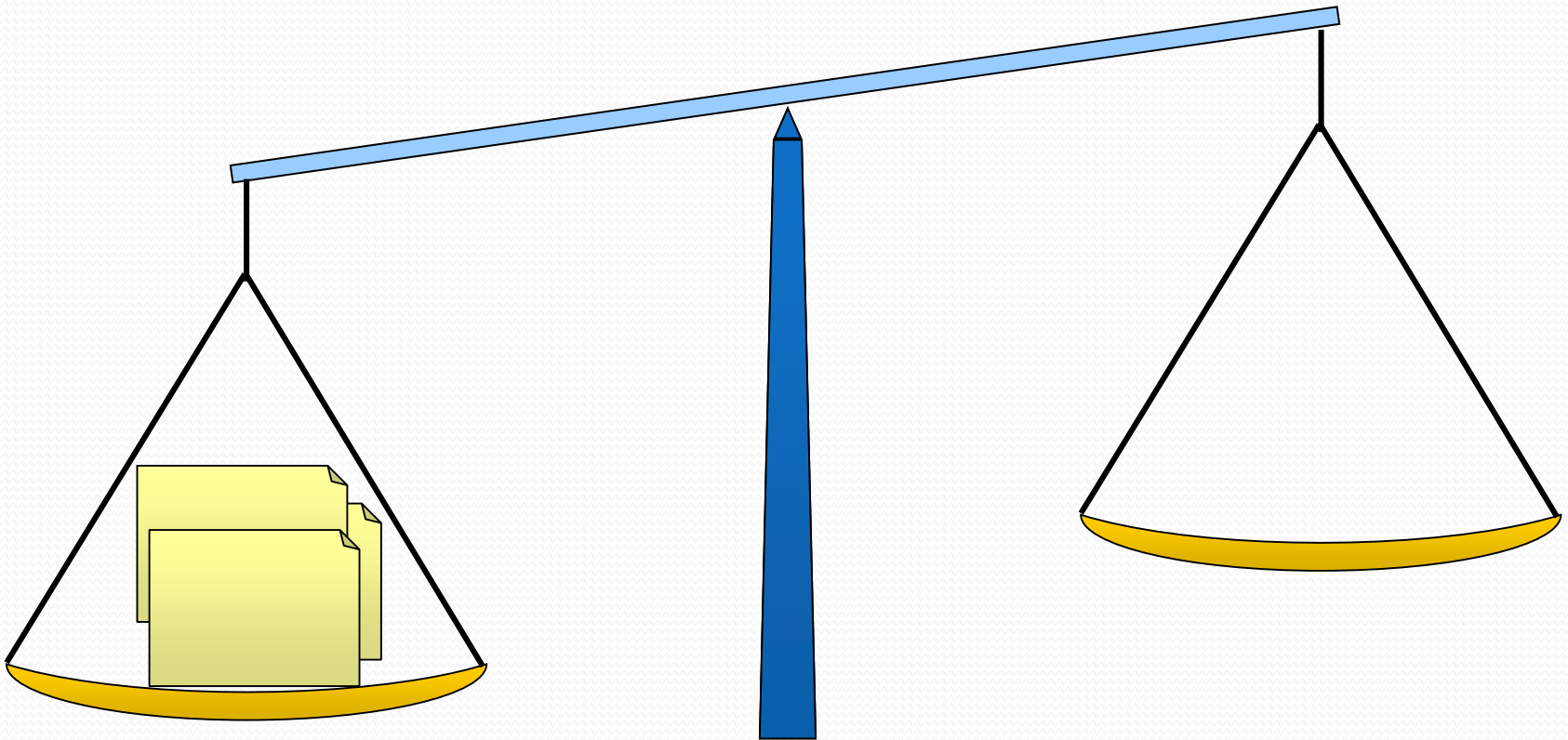
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Background:	Columbia (Col)	Region description:	(transcriptional fusion) Columbia (Col) Genotype: ztl-105 line(4*F2;4*H;) ztl-105; Growth cond: 22C LD Exp cond: 15C RD+RR
Genotype:	ztl-105	update	remove
Marker:	CAB	change all	
Tag type:	Luc		
Data cat.:	Transcriptional fusion		
Lines code:	4*F2;4*H;		
Sample type:	group of seedlings		
Sample origin:	group of seedlings		
Growth stage:	seedling		
Group size:	8		
Growth cond:	22C LD		
Exp cond:	15C RD+RR		
Regions from:	1 to 2		
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	assign		

Getting metadata



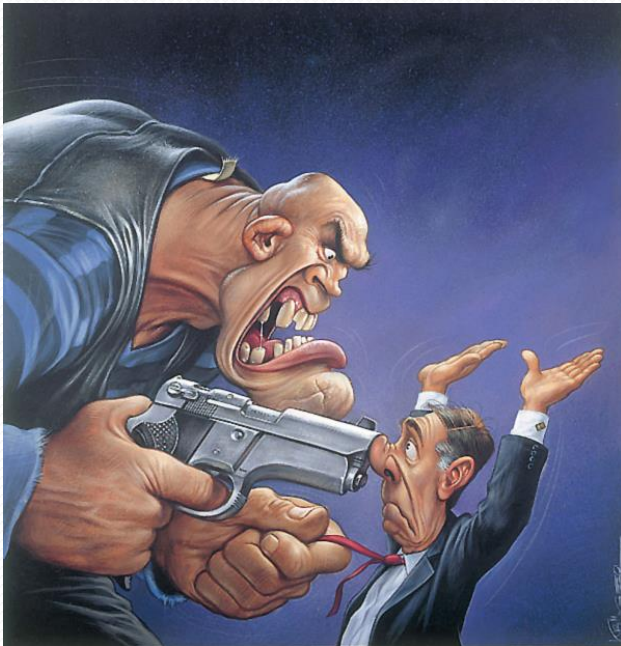
Getting meta data

- No one likes describing data for repositories



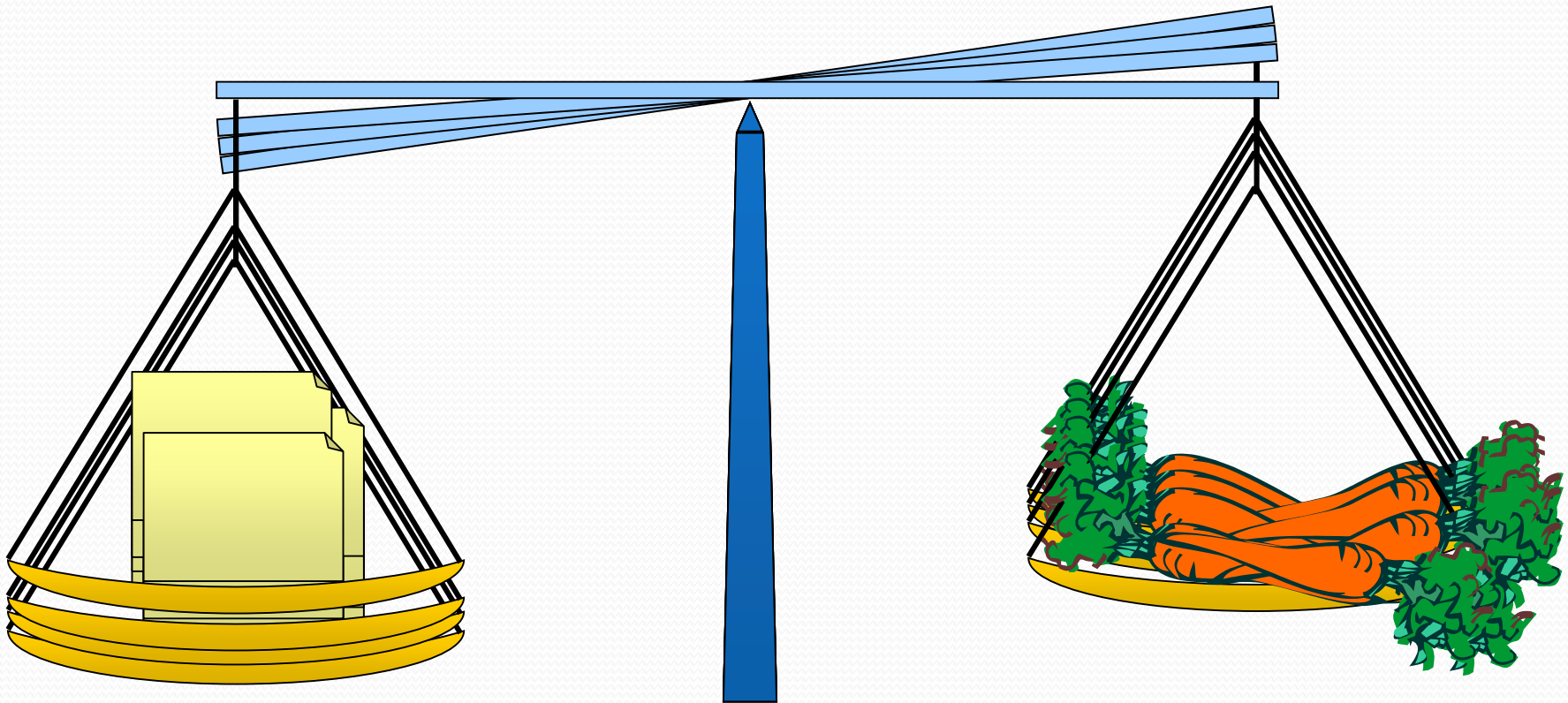
Getting metadata

- No one likes describing data for repositories

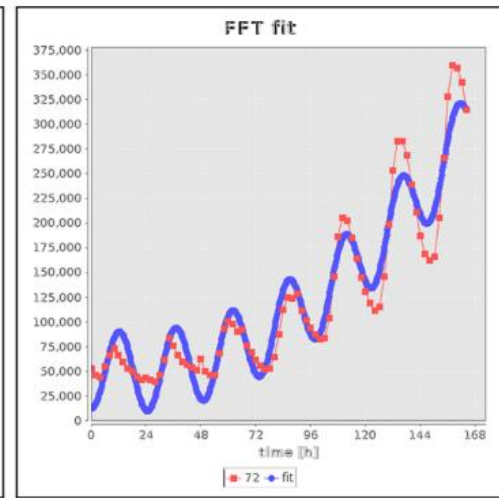
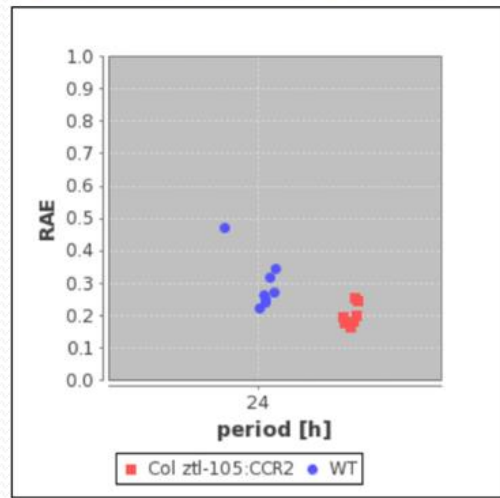
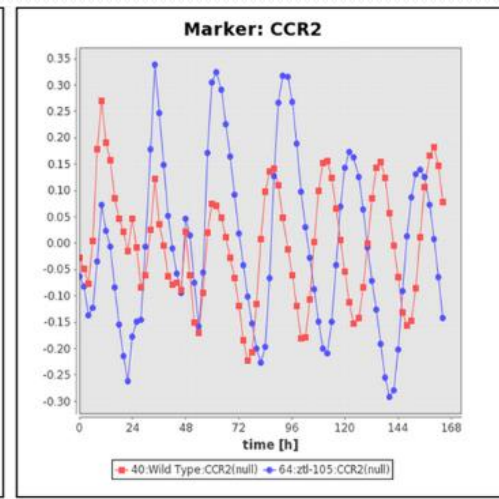
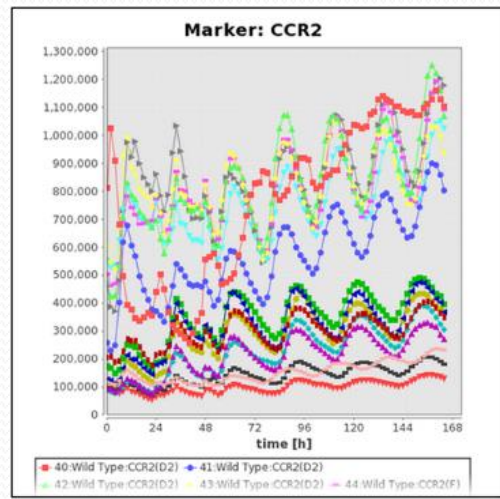


Getting metadata

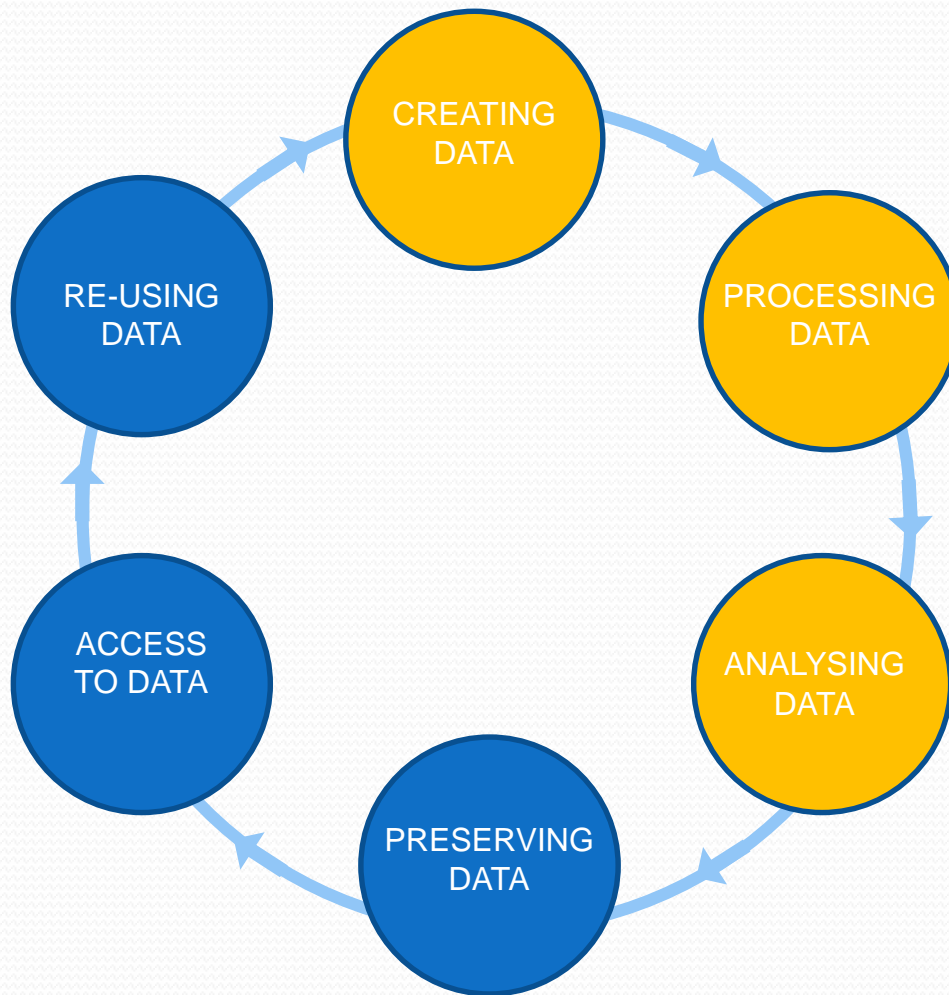
- No one likes describing data for repositories



Tipping the balance

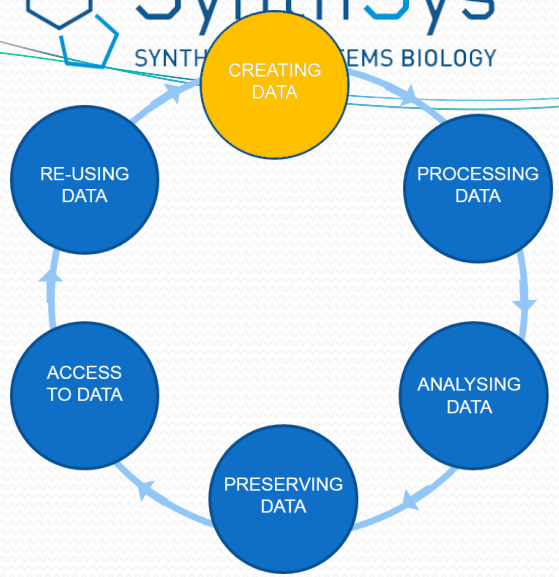


Data management as part of the workflow



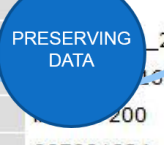
General repositories:
SEEK and OpenBIS

- fine grained metadata,
- logical structure, data relationships
- customizable
- hooks for data processing



Workflow

A1								
	A	B	C	D	E	F	G	H
233	77835s	72	157	481	457	507	458	485
234	78963s	69	156	480	465	507	464	480
235	80091s	77	152	480	462	499	462	475
236	81219s	74	164	502	454	514	464	481
237	82347s	73	152	478	463	510	455	499
238	83475s	76	141	480	465	513	463	495
239	84603s	69	147	499	460	518	450	485
240	85731s	73	146	486	496	492	461	500
241	86859s	77	150	495	491	528	458	500
242	87987s	73	152	507	477	499	478	480
243	89115s	83	151	509	470	531	472	495
244	Date of measurement: 2012-12-16/Time of measurement: 19:20:40							
245	March2012_2EmWL_1gain_M2_80cycles.mth							
246	C:\Users\Public\Documents\Tecan\Magellan\mth\Catie\March2012_2EmWL_1gain_M2_80cy							
247	2012_12_16.wsp							
248	C:\Users\Public\Documents\Tecan\Magellan\wsp\Catie\2012_12_16.wsp							
249	595nm							
250	Unknown user							
251	infinite 200							
252	Instrument serial number: 907001834							
253	Plate							
254	Plate Description: [BD96ft_FluoroBlok] - BD Falcon 96 Flat Transparent/Black							
255	Plate with Cover: Yes							
256	Barcode: No							
257	Part of Plate							
258	Range: A1:H12							
259	Temperature							
260	Mode: On							
261	Temperature: 29.5 °C							
262	Wait for Temperature							
263	Minimum Temperature: 29.0 °C							
264	Maximum Temperature: 30.0 °C							
265	Kinetic Cycle							
266	Number of Cycles: 80							
267	Absorbance							
268	Measurement Wavelength: 595 nm							
269	Measurement Bandwidth: 9 nm							



Workflow

GROUP » GROWTH_RATE » 187992378... Data Set 187992378900 [PS_PLATE_READER]

Data Set Properties

Sample: /PS_GROUP/PL13 [PS_IMG_PLATE]

Measurement: 2012-12-16

Measurement: 19:20:40

Measurement file: ..._2EmWL_1gain_M2_80c...

Wsp file: ..._10.wsp

Instrument: ... 200

Serial Nr: 907001834

Plate desc.: [BD96ft_FluoroBlok] - BD Falcon 96 Flat Transparent/Black

Plate Cell Range: A1:H12

Temperature [C]: 29.5

Min Temp. [C]: 29.0

Max Temp. [C]: 30.0

Shaking: Duration: 1000 sec; Mode: Linear; Amplitude: 6 mm; Frequency: 57.9 rpm;

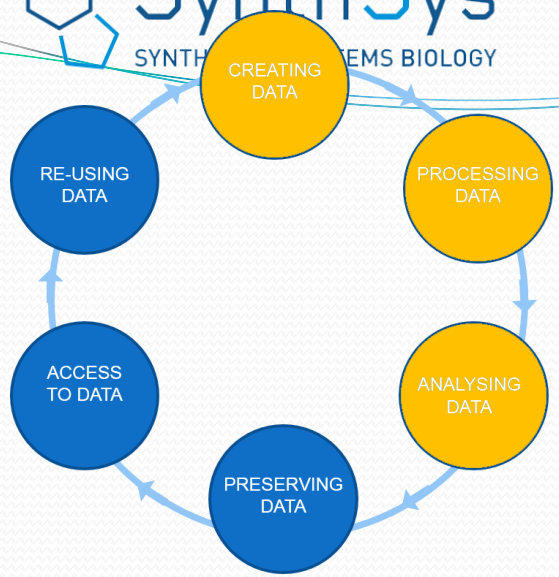
Run Time: 1days 45min 16s

Channels: OD
485-525mGain
485-585mGain

Channel1: OD
Absorbance
595 nm
reads:15

Channel2: 485-525mGain

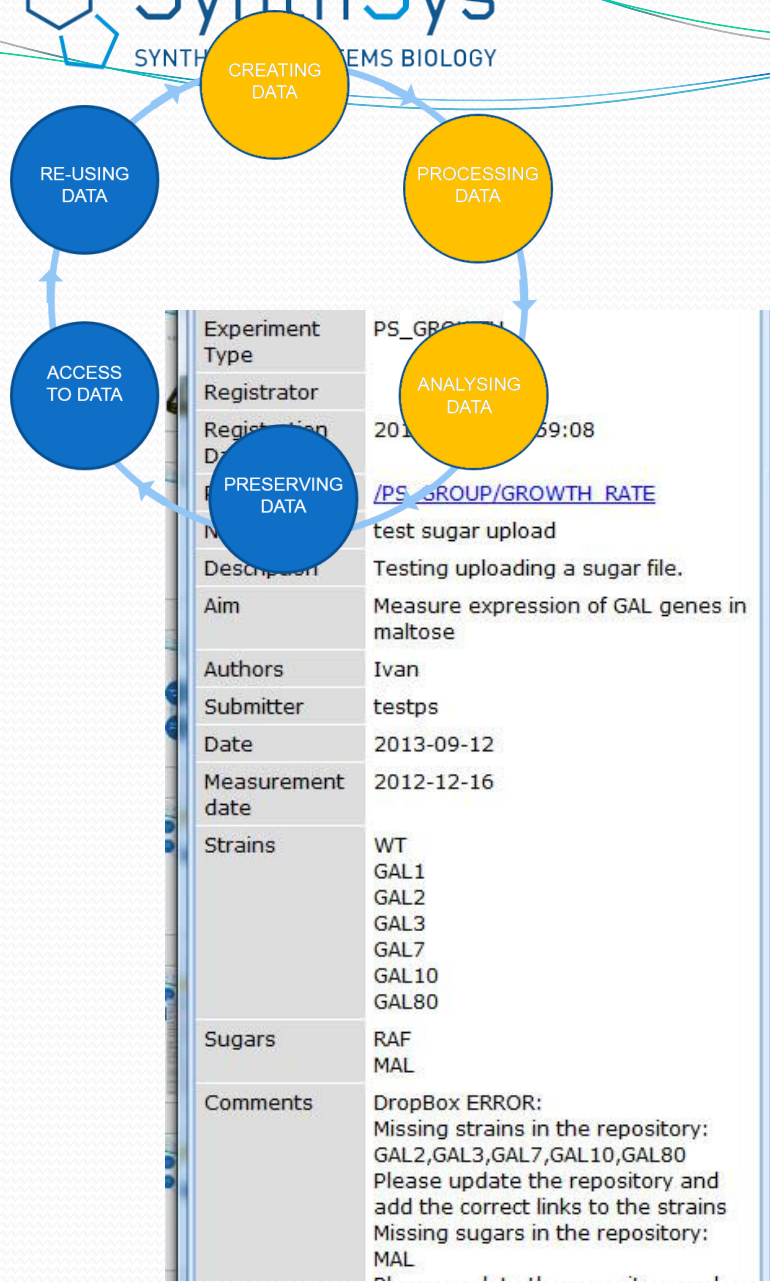
	A	B	C	D	E	F	G	H
233	77835s	72	157	481	457	507	458	485
234	78963s	69	156	480	465	507	464	480
235	80091s	77	152	480	462	499	462	475
236	81219s	74	164	502	454	514	464	481
237	82347s	73	152	478	463	510	455	499
238	83475s	76	141	480	465	513	463	495
239	84603s	69	147	499	460	518	450	485
240	85731s	73	146	486	496	492	461	500
241	86859s	77	150	495	491	528	458	500
242	87987s	73	152	507	477	499	478	480
243	89115s	83	151	509	470	531	472	495
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245	March2012_2EmWL_1gain_M2_80cycles.mth							
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249	595nm							
250	Unknown user							
251	infinite 200							
252	Instrument serial number: 907001834							
253	Plate							
254	Plate Description: [BD96ft_FluoroBlok] - BD Falcon 96 Flat Transparent/Black							
255	Plate with Cover: Yes							
256	Barcode: No							
257	Part of Plate							
258	Range: A1:H12							
259	Temperature							
260	Mode: On							
261	Temperature: 29.5 °C							
262	Wait for Temperature							
263	Minimum Temperature: 29.0 °C							
264	Maximum Temperature: 30.0 °C							
265	Kinetic Cycle							
266	Number of Cycles: 80							
267	Absorbance							
268	Measurement Wavelength: 595 nm							
269	Measurement Bandwidth: 9 nm							
270	...							



Workflow

	A	B	C	D	E	F	G	H
233	77835s	72	157	481	457	507	458	485
234	78963s	69	156	480	465	507	464	480
235	80091s	77	152	480	462	499	462	475
236	81219s	74	164	502	454	514	464	481
237	82347s	73	152	478	463	510	455	499
238	83475s	76	141	480	465	513	463	495
239	84603s	69	147	499	460	518	450	485
240	85731s	73	146	486	496	492	461	500
241	86859s	77	150	495	491	528	458	500
242	87987s	73	152	507	477	499	478	480
243	89115s	83	151	509	470	531	472	495
244	Date of measurement: 2012-12-16/Time of measurement: 19:20:40							
245	March2012_2EmWL_1gain_M2_80cycles.mth							
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	C	D	E	F	G
	2	3	4	5	6
	null in 2% Raf	WT in 2% Raf	WT in 2% Raf	GAL1 in 2% Raf	GAL2 in 2% Raf
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	null in 1% Gal	WT in 1% Gal	WT in 1% Gal	GAL1 in 1% Gal	GAL2 in 1% Gal
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	contaminated	WT in 0.01% Gal	WT in 0.01% Gal	GAL1 in 0.01% Gal	GAL2 in 0.01% Gal
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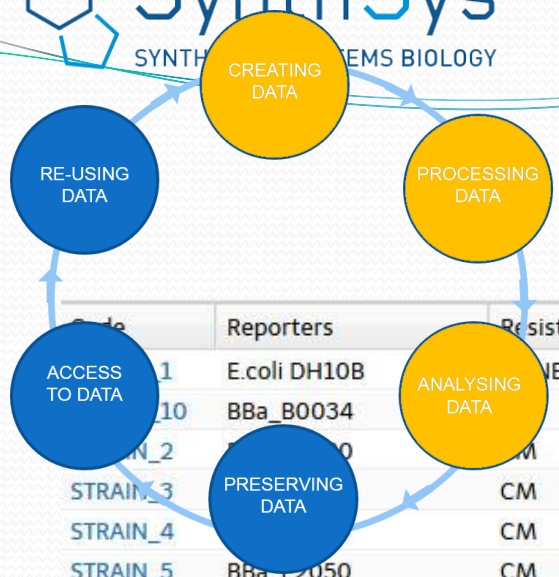
Workflow

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Registrator	
Registration Date	2013-09-12 09:08
Path	/PS_GROUP/GROWTH_RATE
Name	test sugar upload
Description	Testing uploading a sugar file.
Aim	Measure expression of GAL genes in maltose
Authors	Ivan
Submitter	testps
Date	2013-09-12
Measurement date	2012-12-16
Strains	WT GAL1 GAL2 GAL3 GAL7 GAL10 GAL80
Sugars	RAF MAL
Comments	DropBox ERROR: Missing strains in the repository: GAL2,GAL3,GAL7,GAL10,GAL80 Please update the repository and add the correct links to the strains Missing sugars in the repository: MAL

	A	B	C	D	E	F	G	H
233	77835s	72	157	481	457	507	458	485
234	78963s	69	156	480	465	507	464	480
235	80091s	77	152	480	462	499	462	475
236	81219s	74	164	502	454	514	464	481
237	82347s	73	152	478	463	510	455	499
238	83475s	76	141	480	465	513	463	495
239	84603s	69	147	499	460	518	450	485
240	85731s	73	146	486	496	492	461	500
241	86859s	77	150	495	491	528	458	500
242	87987s	73	152	507	477	499	478	480
243	89115s	83	151	509	470	531	472	495
244	Date of measurement: 2012-12-16/Time of measurement: 19:20:40							
245	March2012_2EmWL_1gain_M2_80cycles.mth							
246	C:\Users\Public\Documents\Tecan\Magellan\mth\Catie\March2012_2EmWL_1gain_M2_80cy							
247	2012_12_16.wsp							

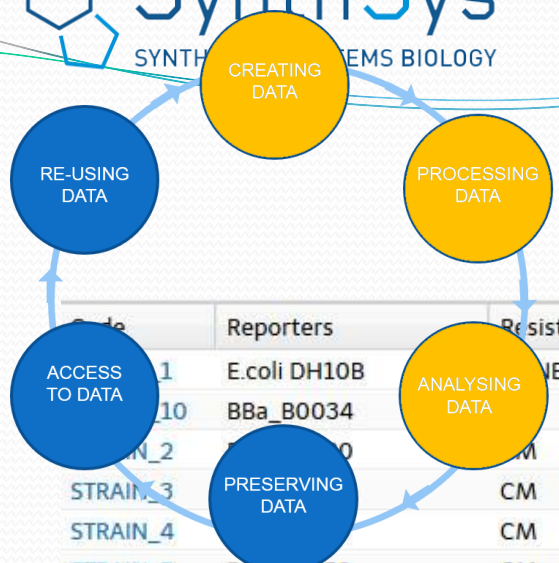
	D	E	F	G
	3	4	5	6
WT in 2% Raf	WT in 2% Raf	GAL1 in 2% Raf	GAL2 in 2% Raf	C
WT in 2% Raf	WT in 2% Raf	GAL1 in 2% Raf	GAL2 in 2% Raf	C
WT in 1% Gal	WT in 1% Gal	GAL1 in 1% Gal	GAL2 in 1% Gal	C
WT in 1% Gal	WT in 1% Gal	GAL1 in 1% Gal	GAL2 in 1% Gal	C
WT in 0.1% Gal	WT in 0.1% Gal	GAL1 in 0.1% Gal	GAL2 in 0.1% Gal	C
WT in 0.1% Gal	WT in 0.1% Gal	contaminated	GAL2 in 0.1% Gal	C
WT in 0.01% Gal	WT in 0.01% Gal	GAL1 in 0.01% Gal	GAL2 in 0.01% Gal	C
WT in 0.01% Gal	WT in 0.01% Gal	GAL1 in 0.01% Gal	GAL2 in 0.01% Gal	C

Workflow

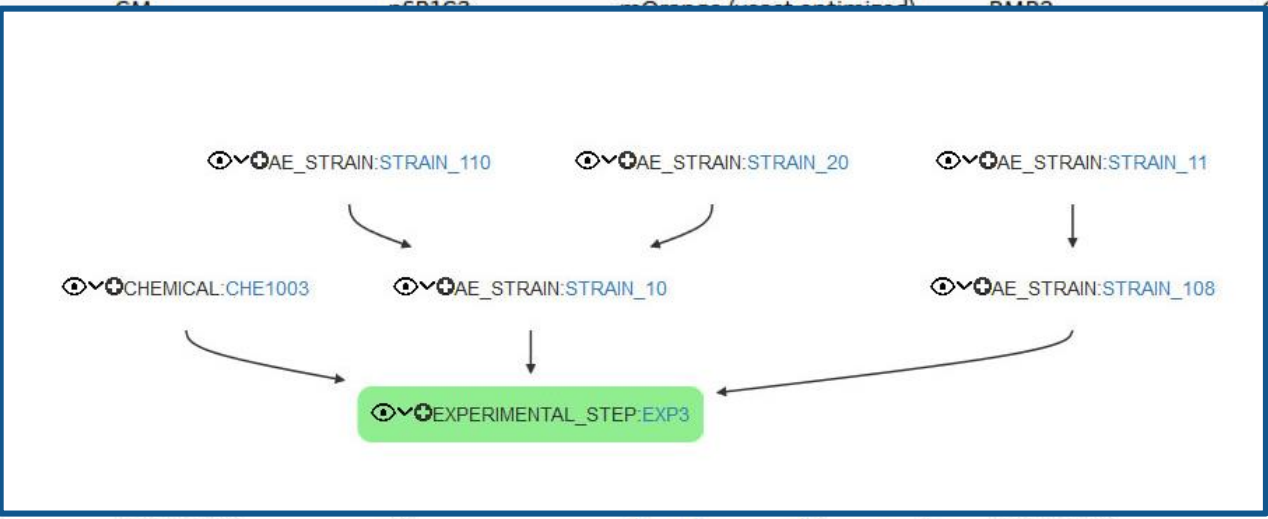


Strain	Reporters	Resistance	Backbone	Description	ORI of Rep	Comments
STRAIN_1	E.coli DH10B	NE	no vectors		NA	Standard E.
STRAIN_10	BBa_B0034	AP	pSB1A2	Strong RBS (tctagaGAAAGAGGAC	COLE1	strong 100%
STRAIN_2	BBa_B0034	AP	pSB1C3	EYFP	PMB2	enhanced ye
STRAIN_3	BBa_B0034	AP	pSB1C3	ECFP	PMB2	enhanced cy
STRAIN_4	BBa_B0034	AP	pSB1C3	mRFP1	PMB1	monomeric
STRAIN_5	BBa_L2050	CM	pSB1C3	mOrange (yeast optimized)	PMB2	Orange fluo
STRAIN_6	BBa_E0040	AP	pSB1A2	GFPmut3b	COLE1	Green fluore
STRAIN_7	BBa_J06702	AP	pSB1A2	mCherry (yeast version also)	COLE1	B0034 RBS/i
STRAIN_8	BBa_J23114	AP	pJ61002	Weak promoter (tttatggctagctca	COLE1	weak promc
STRAIN_9	BBa_J23100	AP	pJ61002	Strong promoter (ttgacggctagct	COLE1	strong promr
STRAIN_11	BBa_J61101	AP	pSB1A2	weak RBS (tctagaGAAAGACAGGA	COLE1	weak 12%
STRAIN_12	E. coli cc118 Lambda ...	NONE	no vectors	cloning strain to maintain RK6 pl	NA	
STRAIN_13	P. putida KT2440	NONE	no vectors	standard P. putida strain without	NA	
STRAIN_14	HB101 RK600	CM	no vectors	E. coli HB101 helper strain to mo	NA	
STRAIN_15	pBBFLP	TC	pBBFLP	Plasmid source Flipase	UNKNOWN	
STRAIN_16	pSEVA 121	AP	pSEVA 121	low copy broad host range plasr	RK2	
STRAIN_17	pSEVA 221	KM	pSEVA 221	low copy broad host range plasr	RK2	
STRAIN_18	pSEVA 321	CM	pSEVA 321	low copy broad host range plasr	RK2	
STRAIN_19	Magnetospirillum ma...	UNKNOWN	NA	glycerol prepared fromsample cu	UNKNOWN	
STRAIN_20	pSEVA 521	TC	pSEVA 521	low copy broad host range plasr	RK2	
STRAIN_21	pSEVA 621	GM	pSEVA 621	low copy broad host range plasr	RK2	
STRAIN_22	pUC18NotI	AP	pUC18NotI	high copy shuttle vector for E. co	UNKNOWN	

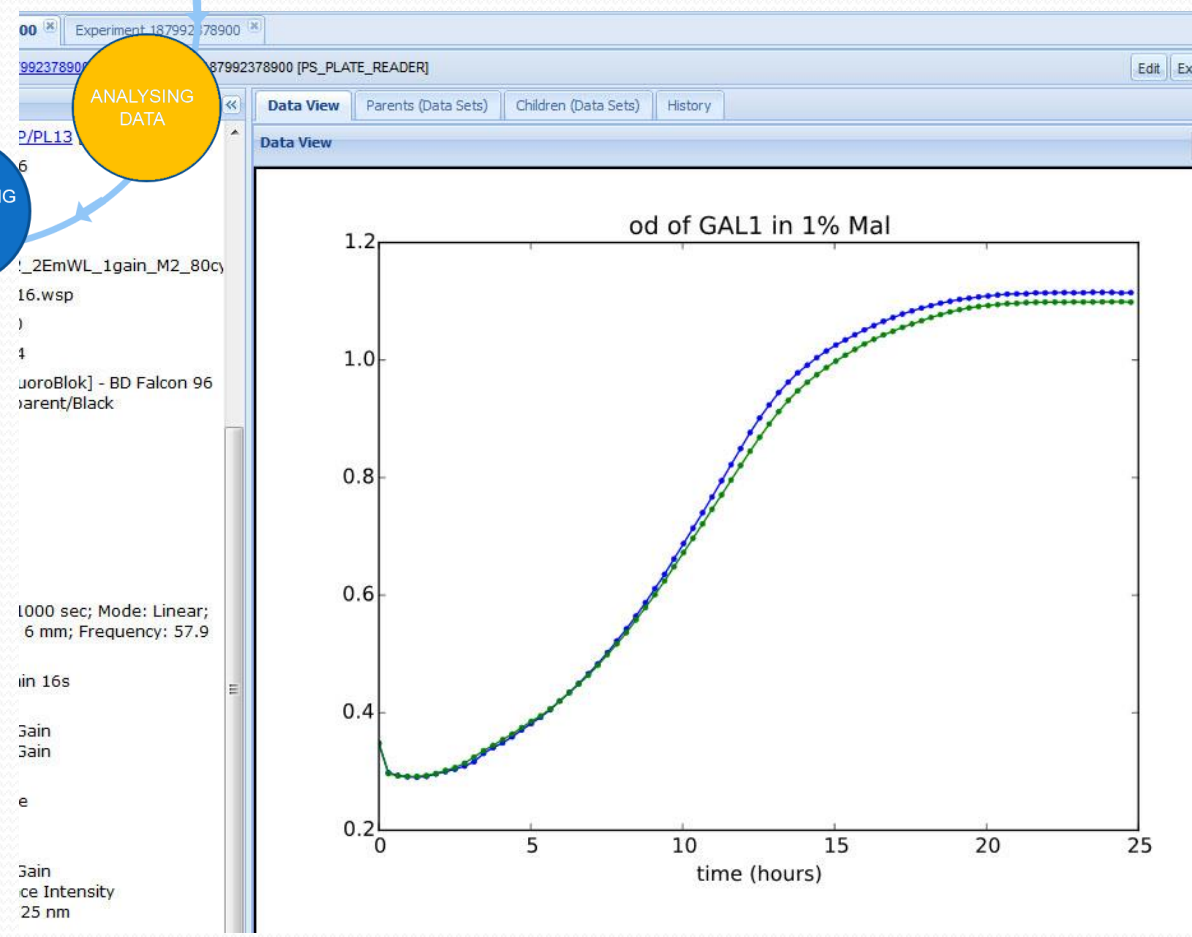
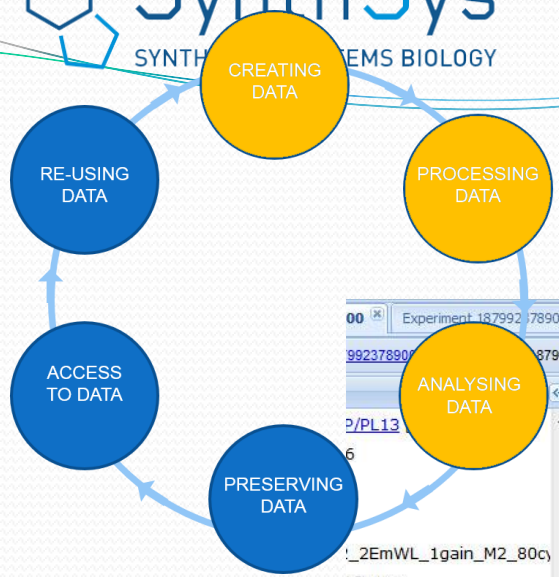
Workflow



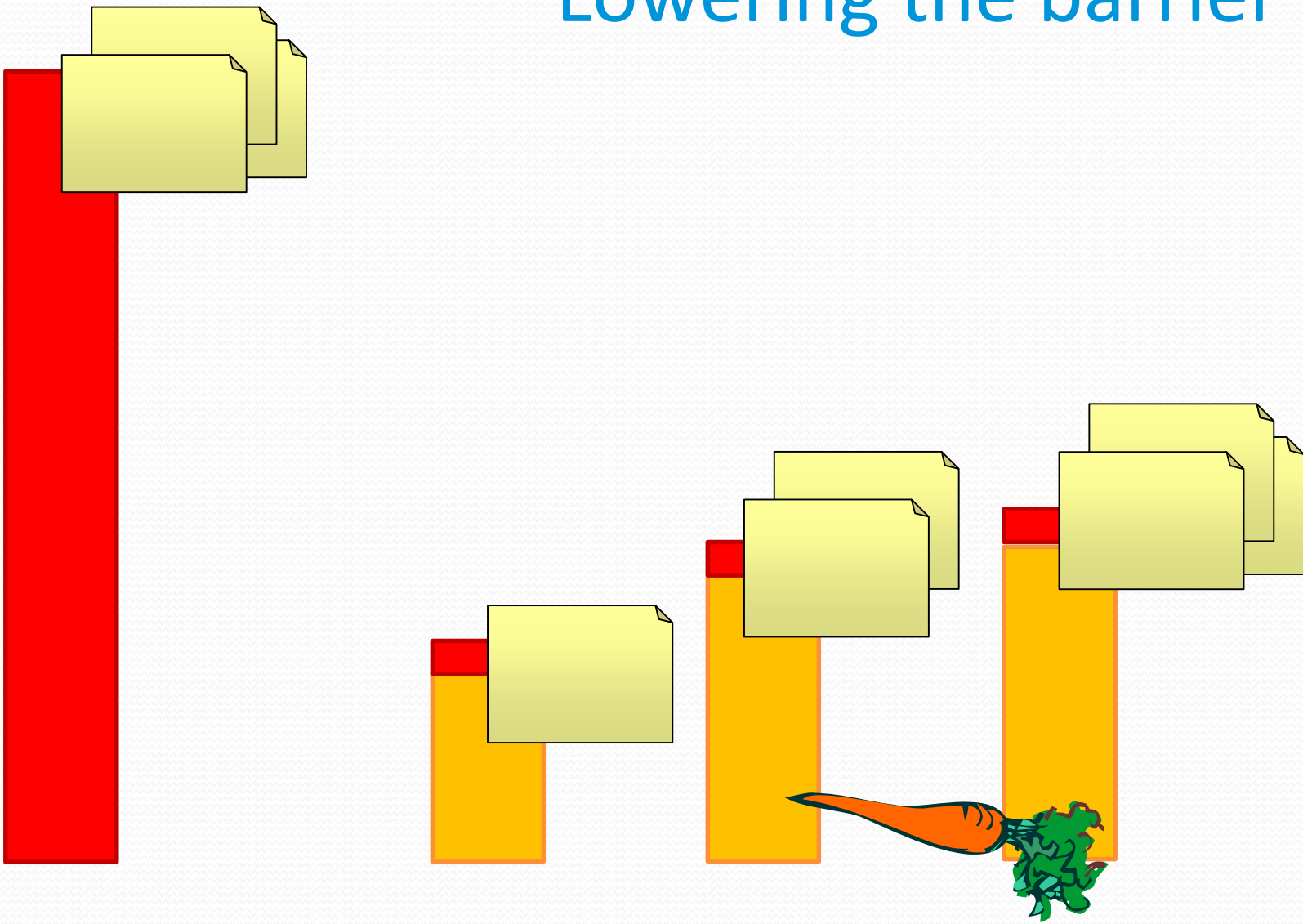
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STRAIN_6	BBa_E0040	CM	pSB1C3	mCherry	PMB2	Green fluore
STRAIN_7	BBa_J06702	CM	pSB1C3	BBa_B0034 RBS/	PMB2	weak promc
STRAIN_8	BBa_J23114	CM	pSB1C3	BBa_J23100	PMB2	strong prom
STRAIN_9	BBa_J23100	CM	pSB1C3	BBa_J61101	PMB2	weak 12%
STRAIN_11	BBa_J61101	CM	pSB1C3		PMB2	
STRAIN_12	E. coli cc118 La	CM	pSB1C3		PMB2	
STRAIN_13	P. putida KT24	CM	pSB1C3		PMB2	
STRAIN_14	HB101 RK600	CM	pSB1C3		PMB2	
STRAIN_15	pBBFLP	CM	pSB1C3		PMB2	
STRAIN_16	pSEVA 121	CM	pSB1C3		PMB2	
STRAIN_17	pSEVA 221	CM	pSB1C3		PMB2	
STRAIN_18	pSEVA 321	CM	pSB1C3		PMB2	
STRAIN_19	Magnetospirillum ma...	UNKNOWN	NA	glycerol prepared from sample cu	UNKNOWN	
STRAIN_20	pSEVA 521	TC	pSEVA 521	low copy broad host range plasr	RK2	
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Workflow



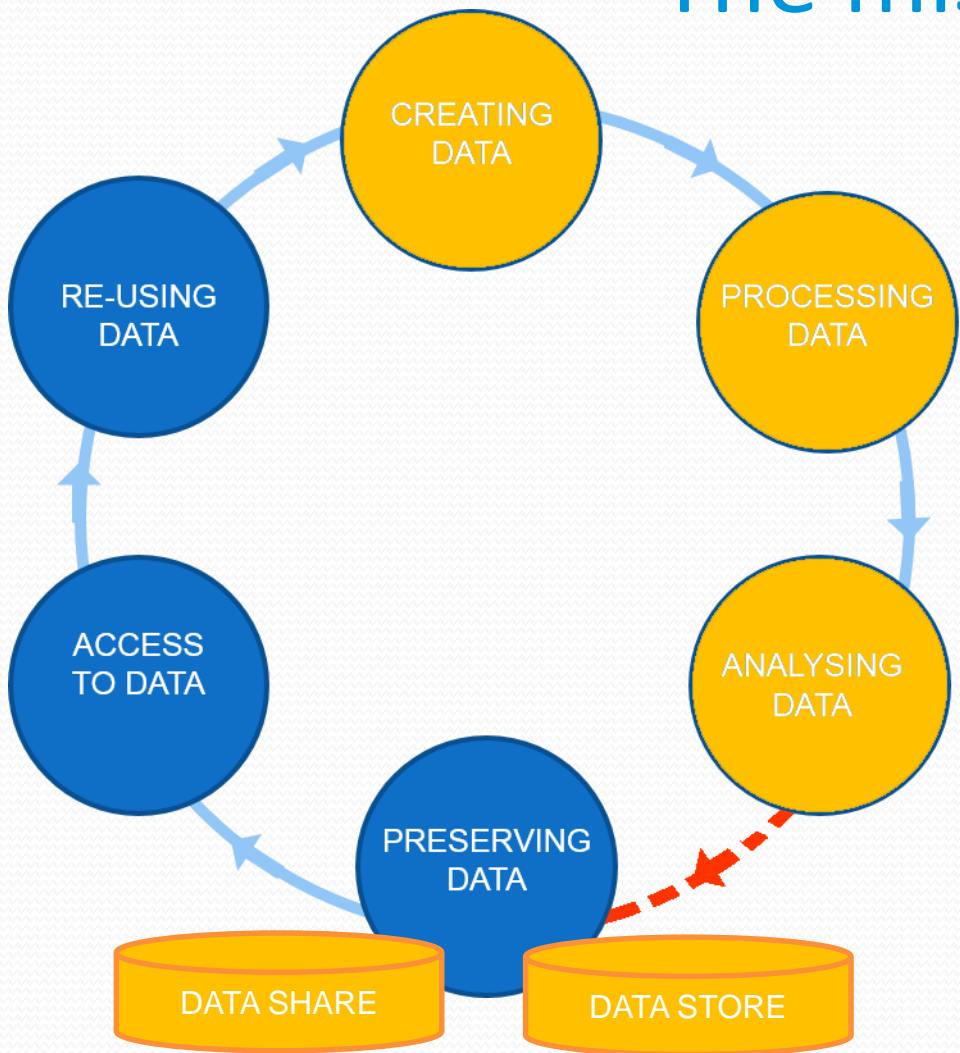
Lowering the barrier



Successful data management?

- is part of the research workflow not an extra burden
- solves user problems not creates new ones
- provides extra value for the user
- focuses on both data consumers and producers

The missing link





SynthSys

SYNTHETIC & SYSTEMS BIOLOGY